

Application of Microbubble Technology to Increase Oxygen Content in The Aquaculture of Tambaqui (*Colossoma macropomum*)

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Abstract

Oxygen plays an important role in aquaculture by serving as an essential component for oxidizing food substances to produce energy, thereby influencing metabolic rates often quantified by oxygen consumption per unit of time. Particularly in intensive aquaculture systems characterized by high stocking densities and maximal feeding rates, oxygen content assumes paramount importance. This study aimed to evaluate the growth performance of tambaqui (Colossoma macropo*mum*) treated with increased oxygen in two different systems. The study was conducted on a laboratory scale by applying Ttest data analysis. The difference in treatment level applied when comparing aeration and microbubble consists of two treatments and three replications. This research used commercial floating pellets PF0 with 25% protein content, the feeding rate was 5% of the biomass. The feed was given daily during the culture for two times a day. Data from the study were analyzed using a T-test on the Statistical Product and Service Solutions (SPSS) software Version 17.0. Results of the study showed that aeration system using microbubble resulted in dissolved oxygen (DO) of 6.5 \pm 0.17, 100% pomfret fish survival rate (SR), Survival Growth Rate (SGR) of 1.83 ± 0.24 , Feed Conversion Ratio (FCR) of 1.37 ± 0.17, Protein Efficiency Ratio (PER) of 2.79 \pm 0.37, Feed Utilization Efficiency (FUE) of 0.67 \pm 0.09, Absolute Weight Growth of 52.02 \pm 1.60; it was better than using aeration. This study has developed a model of tambaqui aquaculture to provide growth value.

INTRODUCTION

Oxygen is one of the most important water quality parameters in aquaculture (Zang *et al.*, 2001; Petranich *et al.*, 2018). Oxygen plays an important role in aquaculture by serving as an essential component for oxidizing food substances to produce energy, thereby influencing metabolic rates often quantified by oxygen

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consumption per unit of time (Pardamean et al., 2021). In intensive aquaculture systems characterized by high stocking densities and maximal feeding rates (Pangestika and Putra, 2020; Jefri et al., 2020), oxygen content is paramount. The heightened stocking density frequently leads to oxygen deficiencies, posing a threat to aquatic life and necessitating a keen consideration of oxygen as a limiting factor (Stickney and Van Liere, 1953). They revealed that lack of oxygen endangers aquatic animals since it causes stress, is susceptible to disease, and can even cause death. The need for oxygen varies depending on the species, size, activity, sex, rate of feed consumption, temperature, and dissolved oxygen concentration (Salvanes et al., 2018).

Dissolved oxygen, a critical aspect of water quality, is intricately linked to respiratory processes crucial for fish survival and well-being (Hasan and Islam, 2020; Hasan and Widodo, 2020; Hasan et al., 2021). Insufficient levels of dissolved oxygen may adversely affect aquaculture water quality, resulting in growth abnormalities and heightened susceptibility to diseases among fish populations (Nafisyah et al., 2018; Tavares-Dias, 2021; Amin et al., 2021; Amin et al., 2022). Mitigating oxygen insufficiency typically involves water changes and aeration; however, the latter often proves inefficient due to challenges in accurately determining the minimum required dissolved oxygen concentration for fish. Consequently, aeration often persists for prolonged periods until oxygen saturation levels are achieved, leading to significant energy wastage, particularly in electrical energy consumption.

A potential solution to this inefficiency lies in the utilization of Microbubble Systems, a cutting-edge aquaculture technology designed to optimize fish cultivation productivity by refining and modifying aquaculture systems. Microbubble technology has garnered substantial interest in recent years owing to its diverse applications across scientific and technological domains. A study by Mahasri *et al.* (2018) demonstrated the efficacy of microbubble technology in augmenting oxygen content in tilapia culture, achieving a concentration of 0.61 pp/minute. Notably, the microbubble model proposed by Luo exhibited superior oxygen distribution compared to conventional high bubble distribution trends (Ren *et al.*, 2019).

Furthermore, microbubble technology has been shown to enhance growth outcomes in seawater fish compared to freshwater species (Duangjai and Punroob, 2017). Microbubble technology is pivotal in enhancing fish survival and growth rates by optimizing water quality through increased dissolved oxygen concentrations. Additionally, microbubbles contribute to waste treatment in culture containers by supplying dissolved oxygen for microbial decomposition of organic and inorganic matter. Due to the lack of studies concerning the application of microbubble technology in Tambaqui cultivation, this study aims to evaluate the comparative effectiveness of aeration versus microbubble technology in enhancing dissolved oxygen content and growth performance of tambaqui.

METHODOLOGY Ethical Approval

All research procedures are carried out according to standards without damaging or polluting the environment and testing laboratory.

Place and Time

The study was conducted from January - December 2020. Comparative biological testing of the use of microbubbles and aeration on the oxygen content and growth of pomfret was conducted at the Fisheries and Marine Sciences Faculty Research Laboratory, Jenderal Soedirman University. This study compared two cultivation system methods aeration and microbubble. Biological tests were conducted by pomfret samples on the growth performance, FCR, PER, and survival of the fish. Maintenance activities were conducted for 30 days with sampling every seven days.

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Research Materials

The tools used in this study were aquarium 40 x 30 x 30, an aerator pump (LP 100), a microbubble pump, a DO meter (Lutron DO-5510), a thermometer, a pH meter digital (00-14.00 pH), an aerator hose, a digital scale, a ruler, and a timer socket. Furthermore, this experiment used pomfret fish (size of 6-7 g) derived from Rempoa Area, Banyumas, Central Java.

Research Design

This research was carried out in a laboratory setting utilizing T-test data analysis. The comparison between aeration and microbubble was conducted using two treatments and three replications. The treatments given were as follows: P1 (The use of aeration) and P2 (The use of microbubble).

Work Procedure Container Preparation

The process of preparing the containers commenced with the cleaning of six units of aquariums with the size of 60 cm \times 35 cm \times 30 cm each. Subsequently, 10L of water was added to each container. The aquariums were thoroughly cleaned, and water filling was carried out over three days to stabilize the water conditions in each aquarium to provide the natural habitat of pomfret fish. The temperature of the research room was maintained within the range of 25-27°C. After the fish exhibited signs of health and activity, they were allowed a three-day period to adapt to their new environment. This adaptation period was crucial to ensure an accurate assessment of the effects of aeration and microbubble cultivation systems.

Preparation and Feeding

Pomfret fish were fed with commercial floating pellets PF0 containing 25% protein, at a feeding rate equivalent to 3% of the

biomass. The fish were fed twice daily during the rearing period. The amount of feed provided was determined based on the weekly weight gain of the sample fish, with adjustments made every seven days (Hossain *et al.*, 2010).

Water Quality Observation

The parameters of the water quality observed in this experiment were dissolved oxygen (DO), temperature, and pH. Dissolved oxygen and temperature were measured using a Lutron DO meter, while pH levels were assessed with a pH meter. Water quality observations were conducted twice daily, in the morning and afternoon, throughout the study.

Data Collection

Collected data included dissolved oxygen (DO) levels, survival rate (SR), Specific Growth Rate (SGR), Feed Conversion Ratio (FCR), Protein Efficiency Ratio (PER), Feed Utilization Efficiency (FUE), and Absolute Weight Growth. (Hossain *et al.*, 2010).

Data Analysis

Data analysis in this study performed the T-test method. The research design compared the average growth achieved using an aeration system versus a microbubble system. Statistical analysis of the data was conducted using T-tests within the Statistical Product and Service Solutions (SPSS) software Version 17.0, enabling conclusions to be drawn regarding the research design.

RESULTS AND DISCUSSIONS

According to the growth performance outcomes and observations concerning the effects of aeration and microbubble treatment on pomfret fish growth, the findings are outlined in Table 1 as follows:

Tuble 1. Observation results of the study parameters.		
Parameter	P1	P2
Dissolved oxygen (mg/L)	$5.38 \pm 0.57^{ m b}$	6.5 ± 0.17^{a}
Survival Rate (SR) (%)	100^{a}	100 ^a
Survival Growth Rate/SGR (%/day)	$1.16\pm0.03^{ m b}$	$1.83 \pm 0.24^{\rm a}$
Feed Conversion Ratio (FCR)	$2.27\pm0.14^{\rm b}$	$1.37\pm0.17^{\mathrm{a}}$
Protein Efficiency Ratio (PER)	$1.66 \pm 0.11^{ m b}$	$2.79 \pm 0.37^{\rm a}$
Feed Utilization Efficiency (FUE)	$0.40 \pm 0.02^{\rm b}$	0.67 ± 0.09^{a}
Absolute Weight Growth (W)	$27.15 \pm 3.67^{\text{b}}$	52.02 ± 1.60^{a}

Table 1.Observation results of the study parameters.

Note: All data listed are mean values (n = 3; $\bar{x} \pm$ SD). The numbers followed by different superscript lowercase letters indicate significant differences (P < 0.05).

Dissolved Oxygen

The statistical analysis revealed a significant difference (P<0.05) in dissolved oxygen content between the aeration and microbubble treatment methods. The average dissolved oxygen content in the microbubble treatment was notably higher compared to the aeration treatment, with respective averages of 6.5 \pm 0.17 mg/L and 5.38 \pm 0.57 mg/L. This study establishes that the microbubble technique offers a more substantial supply of dissolved oxygen than the aeration technique, aligning with findings by Ushikubo et al. (2010) indicating that microbubble systems maintain stable oxygen levels in water, thereby extending the availability of oxygen and increasing dissolved oxygen content. The solubility of oxygen in water is influenced by factors such as water temperature, salinity, agitation, and air pressure.

Additionally, Boyd (1990) noted that oxygen solubility in water decreases with rising temperatures, while gas solubility decreases as salinity levels increase up to 40%, with a salinity value of 6.56 mg/L. When dissolved oxygen concentration falls below saturation levels, oxygen from the atmosphere dissolves into the water, and the process of oxygen dissolution or release is accelerated by the oxygen concentration in air and water (Hepher and Pruginin, 1981). The oxygen dissolves into water through atmospheric diffusion and photosynthesis (Stickney, 1979), primarily through direct diffusion between air and water surfaces (Spotte, 1979). The rate of diffusion is largely contingent on three key factors: oxygen deficiency in water, water surface area exposed to air, and the degree of turbulence (Boyd, 1982).

In addition to its application in fish cultivation, microbubble technology has proven successful in microbiology and plant cultivation. Ago et al. (2005) conducted a microbubble design study focusing on bacterial and plant growth, while Ikeura et al. (2017) explored its impact on plants. Microbubble technology has also been utilized in waste treatment, as demonstrated by Wen et al. (2011). The development of microbubble venturi design methods has enhanced Nile tilapia cultivation and emerged as a viable technology in intensive aquaculture. Microbubbles are engineered with small exhaust air, resulting in highly pressurized air bubbles. This design feature enables microbubbles to expedite the decomposition of organic matter in aquaculture waters, as indicated by Budhijanto et al. (2017).

While microbubble technology has advanced to address oxygen quality in aquaculture waters, its drawback lies in the considerable costs associated with pumps used for oxygen diffusion. Attention must be paid to the appropriate duration of microbubble use to optimize their effectiveness in maintaining scheduled oxygen levels, as suggested by Rofik et al. (2020), who reported an increase in oxygen content of up to 8.8 mg/L within an hour. The oxygen enhancement achieved is contingent upon factors such as water depth, microbubble placement, and resulting water pressure, as detailed by Endo et al. (2008), who observed an increase in oxygen content from 0.52 mg/L to 0.87 mg/L and a decrease in temperature from 0.08-0.12 °C

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through microbubble application in freshwater fish culture.

Survival Rate (SR)

Statistical analyses revealed no significant differences in fish SR between aeration and microbubble treatments (P > 0.05). Both methods exhibited an average SR of 100% for pomfret fish, indicating their robust resistance and adaptability to consistent water quality conditions. Contrary to the assertions of Duangjai and Punroob (2017) regarding microbubble technology's role in enhancing fish survival and growth through water quality maintenance, the findings suggest that the similar SR in both treatments may stem from meeting the optimal dissolved oxygen standards for pomfret fish cultivation.

This aligns with Retnani and Abdulgani's (2013) observations that pomfret fish can thrive even at low salinity levels around 4 ppt, with a survival rate of 100%. In a study by Ayuningrum et al. (2020), microbubble technology applied in intensive vannamei shrimp (Litopenaues vannamei) farming significantly improved SR to approximately 60-100%, whereas non-microbubble treatments exhibited considerably lower SR of around 39-78%. Similarly, catfish farming recorded a high SR of 97.22% (Hidayat et al., 2016). Microbubbles, characterized by small sizes ranging from 50-100 m, facilitate enhanced oxygen diffusion, which accelerates organic matter decomposition and reduces ammonia content in the water, as noted by Deendarlianto et al. (2015) and Jainontee et al. (2019).

Specific Growth Rate (SGR)

The statistical analysis results revealed significant differences in the impact of treatment methods, specifically between aeration and microbubble techniques, on the SGR (P < 0.05). The SGR mean value under the microbubble technique was notably higher compared to that of the aeration method; specifically, the SGR for the microbubble technique stood at $1.83 \pm 0.24\%$ per day, while the aeration technique

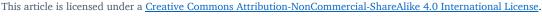
registered $1.16 \pm 0.03\%$ per day. This finding aligns with previous research by Duangjai and Punroob (2017), which highlighted the enhanced growth of pomfret fish through microbubble utilization, particularly in seawater environments.

The variance in pomfret growth rates between these two techniques is attributed to several factors, as noted by Hasan et al. (2015), who mentioned that fish tend to reduce food consumption in low-dissolved oxvgen conditions, thereby affecting growth inversely. Optimal dissolved oxygen concentrations stimulate fish appetite, consequently bolstering growth rates. This elucidates the observed growth rate disparity in pomfret fish between aeration and microbubble techniques. Herivati et al. (2020) further elaborated that microbubble technology facilitated individual fish weight gains and increased fish biomass by 268%. Although aeration improves dissolved oxygen (DO) levels, incorporating microbubble aeration enhances DO levels, growth rates, and fish biomass.

The decline in oxygen availability within the aeration system is considered the primary factor influencing food intake determinants. Reduced oxygen levels result in diminished appetite and growth performance in pomfret fish during feeding sessions, highlighting the importance of adequate oxygen content. Similar findings have been reported in other studies, such as those Oncorhynchus mykiss by Pedersen on (1987); Glencross (2009); D. labrax L (Thetmeyer et al., 1999), Scophthalmus maximus (Pichavant et al., 2000), Anarhichas minor Olafsen (Foss et al., 2003), Ictalurus punctatus (Buentello et al., 2000), O. niloticus (Tran-Duy et al., 2008), and Morone saxatilis (Brandt et al., 2009).

Feed Conversion Ratio (FCR)

Statistical analyses revealed significant differences in treatment methods, specifically between aeration and microbubble techniques, regarding the Feed Conversion Ratio (FCR) (P <0.05). The FCR average calculated for the microbubble treatment



was 1.37 ± 0.17 , whereas that of the aeration treatment was 2.27 ± 0.14 . This suggests that pomfret fish cultured using the microbubble method exhibited improved feed intake compared to those cultured using aeration. These findings align with a study by Heriyati *et al.* (2020), which compared conventional aeration and microbubble techniques in red tilapia aquaculture. In that study, the FCR for the microbubble method was 1.33, while that of the conventional aeration or microbubble had an FCR of 4.42.

The microbubble treatment demonstrated a low FCR value, indicating efficient use of feed by the fish for growth, as evidenced by the high biomass of fish observed at the study's conclusion. The relatively stable Dissolved Oxygen (DO) concentration in the microbubble treatment, ranging from 6.2 to 6.7 mg/L until the study's end, influenced the fish's appetite, metabolism, and growth. This adherence to standard DO values for tilapia culture, set at >3 ppm according to SNI (2009), supports continuous growth. Several studies have reported that microbubbles may enhance fish growth (Onari et al., 2002; Saputra et al., 2018) and impact survival rates (SR). Notably, the SR of the aeration treatment was higher than that of the control (P < 0.05).

The oxygen content in water significantly affects feed conversion; an oxygen content level between 80-120% tends to result in a favorable feed conversion effect. This underscores the importance of adequate oxygen content for nutrient absorption in fish, thereby influencing feed conversion. Oxygen also plays a crucial role as a growth-limiting factor for fish (Welker et al., 2019). Insufficient oxygen levels hinder the protein synthesis of pomfret, as demonstrated by Smith et al. (1996), who observed over a 56% reduction in protein synthesis rates in the liver, red muscle, and white muscle of crucian carp exposed to 48 hours of anoxia.

Feed Utilization Efficiency (FUE)

The statistical analysis results indicated a significant difference (P<0.05) in feed efficiency between the aeration and microbubble treatment methods. The mean feed efficiency in the aeration group was 0.40 ± 0.02 , whereas in the microbubble group, it was 0.67 ± 0.09 . This finding suggests that pomfret fish subjected to microbubble treatment exhibited greater feed utilization efficiency compared to those under aeration treatment.

The feed efficiency achieved in the microbubble treatment reached an optimal level, consistent with Febrianti et al. (2016), who noted that a fine feed efficiency value should exceed 50% or 0.5. This contrasts with the aeration treatment, which did not attain the desired level of feed efficiency, as indicated by Maskur (2005), who linked reduced dissolved oxygen levels in water to decreased feed efficiency. Adequate oxygen levels are crucial for supporting fish growth as they positively influence feed efficiency. Recommended oxygen levels typically range between 60-80%, and sometimes up to 100%, offering various benefits to fish farmers. Higher oxygen content in the culture environment stimulates feed consumption in fish, ultimately promoting favorable growth outcomes, as emphasized by Buentello et al. (2000).

Protein Efficiency Ratio (PER)

Statistical analysis revealed significant differences (P<0.05) in the Protein Efficiency Ratio (PER) between aeration and microbubble treatments for pomfret fish. The average PER value in the aeration group was 1.66 ± 0.11 , whereas in the microbubble group, it was 2.79 \pm 0.37. This discrepancy could be attributed to the dissolved oxygen levels in the water, which impact the activity of protease enzymes. As a result, tambagui were able to efficiently utilize protein-rich food intakes, leading to enhanced PER values. This phenomenon aligns with findings by Mulyani et al. (2014), indicating that increased enzyme activity likely corresponds fish to

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intensifying their efforts to digest nutrients, particularly proteins, to optimize feed protein utilization for growth.

The lower PER value observed in the aeration treatment is attributed to the diminished activity of protease enzymes compared to the microbubble treatment, thereby resulting in Treatment 1 exhibiting a lower PER value than Treatment 2. Additionally, oxygen levels significantly influence antioxidant enzyme activity and malondialdehyde levels in both liver and serum. As noted in previous observations, optimal dissolved oxygen (DO) levels contribute to improved growth performance, feed utilization, and antioxidant response in fish, as highlighted by Yang et al. (2015). Moreover, Abdel-Tawwab et al. (2015) elaborated that low oxygen content in fish culture compromises fish immunity, rendering them more susceptible to diseases. It's noteworthy that providing fish with an oxygen content of 3.5 mg/L leads to decreased growth, while an oxygen content of 6 mg/L positively impacts weight gain, protein retention, amino acids, and feed efficiency, as discussed by Gan et al. (2013). Thus, inadequate oxygen levels pose risks to fish health and may even cause organ damage.

Absolute Weight Growth (W)

The statistical analysis revealed a notable and significant difference (P < 0.05) in pomfret biomass between the aeration and microbubble treatments. The average weight gain in the aeration group was 27.15 \pm 3.67 grams, while in the microbubble group, it was 52.02 ± 1.60 grams. This data indicates that the pomfret biomass was higher in the microbubble treatment compared to aeration. This finding aligns with Herivati et al. (2020), who reported a significant difference (p < 0.05) in red tilapia biomass at the end of their study, with the microbubble treatment showing a 32.5% increase (6.15 kg) compared to conventional aeration treatment (4.64 kg). The consistent oxygen supply from the microbubble contributes aerator to enhanced fish growth as dissolved oxygen is vital for respiration, metabolism, and substance exchange, thereby fueling growth (Boyd, 1982).

This suggests that microbubble treatment can lead to increased fish growth compared conventional aeration. to Budhijanto et al. (2017) similarly applied microbubble aeration in tilapia culture, resulting in accelerated organic matter degradation and subsequent fish growth. Fish intake and growth are notably affected by high oxygen levels compared to low levels. The oxygen needs of fish are also influenced by their size, with smaller fish exhibiting higher feed intake and growth rates than larger fish, especially under low oxygen conditions where hematological adjustments are limited due to a restricted gill surface area (Tran-Duy et al., 2008).

CONCLUSION

Aeration system using microbubble resulted in dissolved oxygen (DO) of $6.5 \pm$ 0.17, 100%pomfret fish survival rate (SR), Survival Growth Rate (SGR) of 1.83 ± 0.24 , Feed Conversion Ratio (FCR) of $1.37 \pm$ 0.17, Protein Efficiency Ratio (PER) of 2.79 \pm 0.37, Feed Utilization Efficiency (FUE) of 0.67 \pm 0.09, Absolute Weight Growth of 52.02 \pm 1.60; it was better than using aeration. This study has developed a model of tambaqui fish aquaculture to provide growth value.

CONFLICT OF INTEREST

There is no conflict of interest in this manuscript between all authors upon writing and publishing this manuscript.

AUTHOR CONTRIBUTION

The contribution of each author is as follows: Ren Fitriadi, Mustika Palupi, and Sesillia Rani Sanudra collected and analyzed data, drafting, and manuscript preparation. Joni Johandra Putra and Muh. Azril participated in the conception and experimental design.

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REFERENCES

- Abdel-Tawwab, М., Hagras, A.E., Elbaghdady, H.A.M. and Monier, M.N., 2015. Effects of dissolved oxygen and fish size on Nile tilapia, Oreochromis niloticus (L.): growth whole-body performance, composition, and innate immunity. Aquaculture International, 23. pp.1261–1274. https://doi.org/10.1007/s10499-015-9882-y
- Ago, K., Nagasawa, K., Takita, J., Itano, R., Morii, N., Matsuda, K. and Takahashi, K., 2005. Development of an aerobic cultivation system by using a microbubble aeration technology. *Journal of Chemical Engineering of Japan*, *38*(9), pp.757–762. https://doi.org/10.1252/jcej.38.757
- Amin, M., Agustono, Ali, M., Prayugo and Hum, N.N.M.F., 2022. Apparent nutrient utilization and metabolic growth rate of Nile tilapia, Oreochromis niloticus, cultured in recirculating aquaculture and biofloc systems. Open Agriculture, 7(1), pp.445-454. https://doi.org/10.1515/opag-2022-0109
- Amin, M., Agustono, Prayugo, Ali, M. and Hum, N.N.M.F., 2021. Comparison of total nutrient recovery in aquaponics and conventional aquaculture systems. *Open Agriculture*, 6(1), pp.682-688. https://doi.org/10.1515/opag-2021-0032
- Ayuningrum, S.B., Istiqomah, I., Rustadi, Triyatmo, B., Isnansetyo, A., Budhijanto, W. and Deendarlianto, 2020. Protective Effect of

Microbubble Aeration and Dietary Probiotics BALSS on Survival and Immunity of White Leg Shrimp (Litopenaeus vannamei) Postlarvae against Acute Low Salinity Stress. Jurnal Perikanan Universitas Gadjah Mada, 22(1), pp.1-8. https://doi.org/10.22146/jfs.51258

Budhijanto, W., Darlianto, D., Pradana, Y.S. and Hartono, M., 2017. Application of micro bubble generator as low cost and high efficient aerator for sustainable fresh water fish farming. *AIP Conference Proceedings*, 1840, 110008.

https://doi.org/10.1063/1.4982338

- Buentello, J.A., Gatlin, D.M. and Neill, W.H., 2000. Effects of water temperature and dissolved oxygen on consumption, daily feed feed utilization and growth of channel catfish (Ictalurus punctatus). Aquaculture, 182(3-4), pp.339-352. https://doi.org/10.1016/S0044-8486(99)00274-4
- Brandt, S.B., Gerken, M., Hartman, K.J. and Demers, E., 2009. Effects of hypoxia on food consumption and growth of juvenile striped bass (Morone saxatilis). Journal of Experimental Marine Biology and Ecology, 381, S143–S149.

https://doi.org/10.1016/j.jembe.200 9.07.028

- Boyd, C.E., 1982. Water Quality Management for Pond Fish Culture. Elsevier, Amsterdam, p.318.
- Boyd, C.E., 1990. Water Quality in Ponds for Aquaculture. Birmingham Publishing Co. Birmingham, Alabama.
- Deendarlianto, Wiratni, Tontowi, A.E., Indarto, I. and Iriawan, A.G.W., 2015. The implementation of a developed microbubble generator on the aerobic wastewater treatment. *International Journal of Technology*, 6(6), pp.924– 930.

https://doi.org/10.14716/ijtech.v6i6 .1696

Duangjai, E. and Punroob, J., 2017. MNB-02 Growth Performance of Asian Sea

Bass (*Lates calcarifer* Bloch) Using Micro Bubbles in Aquaponic System. *Rajamangala University of Technology Lanna*, pp.5-15.

Endo, A., Srithongouthai, S., Nashiki, H., Teshiba, I., Iwasaki, T., Hama, D. and Tsutsumi, H., 2008. DO-increasing effects of a microscopic bubble generating system in a fish farm. *Marine Pollution Bulletin*, *57*(1–5), pp.78–85.

https://doi.org/10.1016/j.marpolbul .2007.10.014

- Febrianti, K., Sukarti., K. and Pebrianto, C.A., 2016. Pengaruh Perbedaan Sumber Asam Lemak pada Pakan Terhadap Pertumbuhan Ikan Bawal Bintang (*Trachinotus blochii*, lecepede). *Aquawarman Jurnal Sains dan Teknologi Akuakultur*, 2(1), pp.24-33. https://sites.google.com/site/jurnala quawarmancom/volume-2-terbit-1april-2016?authuser=0
- Foss, A., Vollen, T. and Øiestad, V., 2003. Growth and oxygen consumption in normal and O₂ supersaturated water, and interactive effects of O₂ saturation and ammonia on growth in spotted wolffish (*Anarhichas minor* Olafsen). *Aquaculture*, 224(1-4), pp.105–116. https://doi.org/10.1016/S0044-8486(03)00209-6
- Gan, L., Liu, Y.J., Tian, L.X., Yue, Y.R., Yang,
 H.J., Liu, F.J., Chen, Y.J. and Liang,
 G.Y., 2013. Effects of dissolved oxygen and dietary lysine levels on growth performance, feed conversion ratio and body composition of grass carp, *Ctenopharyngodon idella*. *Aquaculture Nutrition*, 19(6), pp.860–869.

https://doi.org/10.1111/anu.12030

Glencross, B.D., 2009. Reduced water oxygen levels affect maximal feed intake, but not protein or energy utilization efficiency of rainbow trout (*Oncorhynchus mykiss*). Aquaculture Nutrition, 15(1), pp.1–8. https://doi.org/10.1111/j.1365-2095.2007.00562.x

- Hasan, V., Valen, F.S., Islamy, R.A., Widodo, M.S., Saptadjaja, A.M. and Islam. I.. 2021. Short Communication: presence of the vulnerable freshwater goby Sicyopus auxilimentus (Gobiidae, Sicydiinae) Sangihe Island. Indonesia. on Biodiversitas 22(2),pp.571-579. https://doi.org/10.13057/biodiv/d220208
- Hasan, V. and Islam, I., 2020. First inland record of Bull shark *Carcharhinus leucas* (Müller & Henle, 1839) (Carcharhiniformes: Carcharhinidae) in Celebes, Indonesia. *Ecologica Montenegrina*, 38, pp.12-17. https://doi.org/10.37828/em.2020.3 8.3
- Hasan, V. and Widodo M.S., 2020. Short Communication: The presence of Bull Carcharhinus shark leucas (Elasmobranchii: Carcharhinidae) in fresh the waters of Sumatra, Indonesia. Biodiversitas, 21(9), pp.4433-4439. https://doi.org/10.13057/bio-

div/d210962

- Hasan, Z., Masjamsir and Iskandar, 2015.
 Pemanfaatan Teknologi Aerasi Berbasis Energi Surya Untuk Memperbaiki Kualitas Air dan Meningkatkan Pertumbuhan Ikan Nila Di KJA Waduk Cirata. *Jurnal Akuatika*, 6(1), pp.68-78. http://jurnal.unpad.ac.id/akuatika/article/view/5966
- Hepher, B. and Pruginin, Y., 1981. Commercial Fish Farming: With Special Reference to Fish Culture In Israel. John Wiley and Sons. New York.
- Heriyati, E., Rustadi, Isnansetyo, A. and Triyatmo, B., 2020. Uji Aerasi Microbubble dalam Menentukan Kualitas Air, Nilai Nutrition Value Coefficient (NVC), Faktor Kondisi (K) dan Performa pada Budidaya Nila Merah (*Oreocrhomis* Sp.). Jurnal Pertanian Terpadu, 8(1), pp.27-41. https://doi.org/10.36084/jpt..v8i1.2 32
- Hidayat, K.W., Supriyono, E., Setiyanto, D.D. and, Widiyati, A., 2016. Effect of

Cite this document Fitriadi, R., Palupi, M., Sanudra, S.R., Putra, J.J. and Azril, M., 2024. Application of Microbubble Technology to Increase Oxygen Content in The Aquaculture of Tambaqui (*Colossoma macropomum*). *Journal of Aquaculture and Fish Health*, *13*(3), pp.328-339.

three simple design micro-pore aeration on growth and survival of hybrid catfish Pangasius sp. *International Journal of Fisheries and Aquatic Studies*, 4(4), pp.170–172. https://www.fisheriesjournal.com/ar chives/2016/vol4issue4/PartC/4-3-3-679.pdf

- Hossain, M.A., Almatar, S.M. and James, C.M., 2010. Optimum Dietary Protein Level for Juvenile Silver Pomfret, *Pampus argenteus* (Euphrasen). *Journal Of The World Aquaculture Society*, 41(5), pp.710-720. http://dx.doi.org/10.1111/j.1749-7345.2010.00413.x
- Ikeura, H., Takahashi, H., Kobayashi, F., Sato, M. and Tamaki, M., 2017. Effect of different microbubble generation methods on growth of Japanese mustard spinach. *Journal of Plant Nutrition*, 40(1), pp.115–127. https://doi.org/10.1080/01904167.2 016.1201498
- Jainontee, K., Norarat, R., Boonchuay, S., Thongdon-a, R., Unsing, A., Booncharoen, P., Janwong, W. and Wesanarat, P., 2019. Preliminary study of the effects of air-fine (Micro/ nano) bubbles (fb) on the growth rate of tilapia in phan district, chiang rai, thailand. International Journal of Plasma Environmental Science and Technology, 12(2),pp.84-88. https://doi.org/10.34343/ijpest.201 9.12.02.084
- Jefri, M., Satyantini., W.H., Sahidu, A.M., Nindarwi, D.D. and Rozi, 2020. Application of Probiotics for Organic Matter and Enhancement of Growth Performance in White Shrimp (*Litopenaeus vannamei*). Jurnal Ilmiah Perikanan dan Kelautan, 12(1), pp.97-104.

https://doi.org/10.20473/jipk.v12i1. 16618

Mahasri, G., Saskia, A., Apandi, P.S., Dewi, N.N., Rozi and Usuman, N.M., 2018. Development of an aquaculture system using nanobubble technology for the optimation of dissolved oxygen in culture media for nile tilapia (*Oreochromis niloticus*). *IOP Conference Series: Earth and Environmental Science*, *137*, 012046. https://doi.org/10.1088/1755-

1315/137/1/012046

- Maskur, 2005. Kondisi kualitas lingkungan perairan umum di Jawa Barat. BBAT Sukabumi. p.5.
- Mulyani, Y.S., Yulisman and Fitrani, M., 2014. Pertumbuhan dan Efisiensi Pakan Ikan Nila (*Oreochromis niloticus*) Yang Dipuasakan Secara Periodik. *Jurnal Akuakultur Rawa Indonesia*, 2(1), pp.1-9. https://doi.org/10.36706/jari.v2i1.1 958
- Nafisyah, A.L., Masithah, E.D., Matsuoka, K., Lamid, M., Alamsjah, M.A., Ohara, S. and Koike, K., 2018. Cryptic occurrence of *Chattonella marina* var. *marina* in mangrove sediments in Probolinggo, East Java Province, Indonesia. *Fisheries Science*, *84*, pp.877-887.

https://doi.org/10.1007/s12562-018-1219-0

Onari, H., Maeda, K., Matsuo, K., Yamahara, Y., Watanabe, K. and Ishikawa, N., 2002. Effect of micro-bubble technique on oyster cultivation. *Annual Journal of Hydraulic Engineering*, *46*, pp.1163-1168. https://doi.org/10.2208/prohe.46.1

163

Pangestika, W. and Putra, S., 2020. Fish Feed Formulation with the Addition of Sludge of Dairy Wastewater and Fermented Wheat Bran. *Jurnal Ilmiah Perikanan dan Kelautan*, 12(1), pp.21-30. https://doi.org/10.20473/jipk.v12i1.

18110

Pardamean, M.A., Islamy, R.A., Hasan, V., Herawati, E.Y. and Mutmainnah, N., 2021. Identification and physiological characteristics of potential indigenous bacteria as bioremediation agent in the wastewater of sugar factory. *Sains Malaysiana*, *50*(2), pp.279-286.

Cite this document Fitriadi, R., Palupi, M., Sanudra, S.R., Putra, J.J. and Azril, M., 2024. Application of Microbubble Technology to Increase Oxygen Content in The Aquaculture of Tambaqui (*Colossoma macropomum*). *Journal of Aquaculture and Fish Health*, *13*(3), pp.328-339.

http://dx.doi.org/10.17576/jsm-2021-5002-01

- Pedersen, C.L., 1987. Energy budgets for juvenile rainbow trout at various oxygen concentrations. *Aquaculture*, 62(3-4), pp.289–298. https://doi.org/10.1016/0044-8486(87)90171-2
- Petranich, E., Covelli, S., Acquavita, A., De Vittor, C., Faganeli, J. and Contin, M., 2018. Benthic nutrient cycling at the sediment-water interface in a lagoon fish farming system (northern Adriatic Sea, Italy). Science of the Total Environment, 644, pp.137-149. https://doi.org/10.1016/j.scitotenv.2018.06.310
- Pichavant, K., Person-Le-Ruyet, J., Le Bayon, N., Sévère, A., Le Roux, A., Quéméner, L., Maxime, V., Nonnotte, G. and Boeuf, G., 2000. Effects of hypoxia on growth and metabolism of juvenile turbot. *Aquaculture*, *188*(1-2), pp.103–114. https://doi.org/10.1016/S0044-8486(00)00316-1
- Ren, F., Noda, N.A., Ueda, T., Sano, Y., Takase, Y., Umekage, T., Yonezawa, Y. and Tanaka, H., 2019. CFD-PBM approach for the gas-liquid flow in a nanobubble generator with honeycomb structure. *Journal of Dispersion Science and Technology*, 40(2), pp.306-317.

https://doi.org/10.1080/01932691.2 018.1470009

- Retnani, H.T. and Abdulgani, N., 2013. Pengaruh Salinitas terhadap Kandungan Protein dan Pertumbuhan Ikan Bawal Bintang (*Trachinotus blochii*). Jurnal Sains dan Seni ITS, 2(2), pp.1-6. http://dx.doi.org/10.12962/j233735 20.v2i2.4051
- Rofik, D.A., Kardiman, Sumarjo, H.J. and Noubnome, V., 2020. Perancangan Dan Analisis Alat Microbubble Generator (Mbg). Gorontalo Journal of Infrastructure and Science Engineering, 3(2), pp.24–30.

https://doi.org/10.32662/gojise.v3i2 .1206

- Salvanes, A.G.V., Christiansen, H., Taha, Y., Henseler, C., Seivåg, M.L., Kjesbu, O.S., Folkvord, A., Utne-Palm, A.C., Currie, B., Ekau, W., van der Plas, A.K. and Gibbons, M.J., 2018. Variation in morphology growth. and reproduction of the bearded goby (Sufflogobius bibarbatus) in varying oxygen environments of northern Benguela. Journal of Marine Systems, 188. pp.81-97. https://doi.org/10.1016/j.jmarsys.20 18.04.003
- Saputra, H.K., Nirmala, K., Supriyono, E. and Rochman, N.T., 2018. Micro/ Nano Bubble Technology : Characteristics and Implications Biology Performance of Koi *Cyprinus carpio* in Recirculation Aquaculture System (RAS). *Omni-Akuatika*, 14, pp.29-36. http://dx.doi.org/10.20884/1.oa.20 18.14.2.539
- Smith, R.W., Houlihan. D.F., Nilsson, G.E. and Brechin, J.G., 1996. Tissuespecific changes in protein synthesis rates in vivo during anoxia in crucian carp. *American Journal of Physiology*, 271, pp.R897–R904. https://doi.org/10.1152/ajpregu.1996.271.4.R897
- SNI (Standar Nasional Indonesia), (2009). Produksi Induk Ikan Nila Hitam (Oreochromis niloticus Bleeker) Kelas Induk Pokok. Badan Standarisasi Nasional (BSN). Jakarta.
- Spotte, S., 1979. Sea Water Aquarium. The Captive Environment. John Wiley and Sons. New York-Christer-Brisbane-Toronto.
- Stickney, R.R., 1979. Principles of Warm Water Aquaculture. John Wiley and Sons Inc. New York. pp.223-229.
- Stickney, J.C. and Van Liere, E.J., 1953. Acclimatization to low oxygen tension. *Physiological Reviews*, 33(1), pp.13-34.

https://doi.org/10.1152/physrev.19 53.33.1.13

Cite this document Fitriadi, R., Palupi, M., Sanudra, S.R., Putra, J.J. and Azril, M., 2024. Application of Microbubble Technology to Increase Oxygen Content in The Aquaculture of Tambaqui (*Colossoma macropomum*). *Journal of Aquaculture and Fish Health*, *13*(3), pp.328-339.

Tavares-Dias, M., 2021. Toxic, physiological, histomorphological, growth performance and antiparasitic effects of copper sulphate in fish aquaculture. *Aquaculture*, 535, 736350.

> https://doi.org/10.1016/j.aquaculture.2021.736350

Thetmeyer, H., Waller, U., Black, K.D., Inselmann, S. and Rosenthal, H., 1999. Growth of European sea bass (*Dicentrarchus labrax* L.) under hypoxic and oscillating oxygen conditions. *Aquaculture*, 174(3-4), pp.355–367. https://doi.org/10.1016/S0044-

8486(99)00028-9

- Tran-Duy, A., Schrama, J.W., van Dam, A.A. and Verreth, J.A.J., 2008. Effects of oxygen concentration and body weight on maximum feed intake, growth and hematological parameters of Nile tilapia, *Oreochromis niloticus*. *Aquaculture*, 275(1–4), pp.152–162. https://doi.org/10.1016/j.aquacultur e.2007.12.024
- Ushikubo, F.Y., Furukawa, T., Nakagawa, R., Masatoshi, E., Makino, Y., Kawagoe, Y., Shiina, T. and Oshita, S., 2010. Evidence of the existence and the stability of nano-bubbles in water. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 361(1-3), pp.31-37.

https://doi.org/10.1016/j.colsurfa.2 010.03.005

Welker, T.L., Overturf, K. and Abernathy, J., 2019. Effect of aeration and oxygenation on growth and survival of rainbow trout in a commercial serial-pass, flow-through raceway system. Aquaculture Reports, 14, 100194.

https://doi.org/10.1016/j.aqrep.201 9.100194

Wen, L.H., Ismail, A.B., Menon, P.M., Saththasivam, J., Thu, K. and Choon, N.K., 2011. Case studies of microbubbles in wastewater treatment. *Desalination and Water Treatment*, 30(1–3), pp.10–16. https://doi.org/10.5004/dwt.2011.1 217

- Yang, K., Fan, Q., Zhang, L., Li, B., Gao, Y., Zeng, K., Wang, Q., Zhu, S. and Fang, G., 2015. Effect of dissolved oxygen levels on growth performance, energy budget and antioxidant responses of yellow catfish, Pelteobagrus fulvidraco (Richardson). *Aquaculture Research*, 46(8), pp.2025–2033. https://doi.org/10.1111/are.12359
- Zang, C., Huang, S., Wu M., Du, S., Scholz, M., Gao, F., Guo, Y. and Dong, Y., 2011. Comparison of relationships between pH, dissolved oxygen, and chlorophyll for aquaculture and nonaquaculture waters. *Water, Air, and Soil Pollution, 219*, pp.157-174. https://doi.org/10.1007/s11270-010-0695-3

Cite this document Fitriadi, R., Palupi, M., Sanudra, S.R., Putra, J.J. and Azril, M., 2024. Application of Microbubble Technology to Increase Oxygen Content in The Aquaculture of Tambaqui (*Colossoma macropomum*). *Journal of Aquaculture and Fish Health*, *13*(3), pp.328-339.

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