


Scientific Journal of Fisheries and Marine

JIPK

(JURNAL ILMIAH PERIKANAN DAN KELAUTAN)

Short Communication

Analysis of White Feces Disease (WFD) caused by *Vibrio* sp. and Dinoflagellata in Vannamei Shrimp (*Litopenaeus vannamei*) in Brackishwater Culture Pond

Moh. Awaludin Adam^{1,2}, Irawati Mei Widiastuti³, Ernawati⁴, Achmad Yani Yayan⁵, Era Insivitawati⁶, Yuliana⁷, Rini Fitriarsari Pakaya⁸, Agoes Soegianto⁹, and Ach. Khumaidi^{1*} 

¹Faculty of Science and Technology, University of Ibrahimy, Situbondo, East Java. Indonesia

²Research Center for Limnology, BRIN, Cibinong, Bogor. Indonesia

³Department of Aquaculture, Faculty of Animal Husbandry and Fisheries, University of Tadulako, Palu, Central Sulawesi. Indonesia

⁴Technology of Fisheries Product, Faculty of Agriculture, University of Yudharta, Pasuruan, East Java. Indonesia

⁵Stations of PSDKP Ambon, Marine and Fisheries Ministry. Indonesia

⁶Polytechnics of Marine and Fisheries, KKP, Sidoarjo, East Java. Indonesia

⁷SMKN Keruak 1, East Lombok, NTB. Indonesia

⁸Departement of Marine and Fisheries, Gorontalo. Indonesia

⁹Faculty of Science and Technology, University of Airlangga, Surabaya, East Java. Indonesia



ARTICLE INFO

Received: Juny 05, 2021

Accepted: October 07, 2021

Published: October 19, 2021

*) Corresponding author:

E-mail: ach.khumaidi@gmail.com

Keywords:

Clinical Symptom

Dinoflagellata

TOM

Vibrio sp.

White Feces

This is an open access article under the CC BY-NC-SA license (<https://creativecommons.org/licenses/by-nc-sa/4.0/>)

Abstract

Shrimp disease that currently causes economic loss to shrimp farmers is White Feces Disease (WFD). This disease appeared due to several factors, such as poor pond management, unhealthy shrimp seed, and poor water quality which resulted in the appearance of *Vibrio* sp. bacteria and Dinoflagellata. This study aimed to analyze the cause of WFD outbreak in vannamei shrimp pond. The study method through direct experiment was applied in shrimp pond. Sampling was performed three times in each feeding tray to collect ten shrimps. Overall, sampling was performed twice a week. The Sample Survey Method was used to collect sample in this study. Result of study showed that clinical symptom was observed through changes in pattern and behavior of vannamei shrimp during culture. However, this observation resulted in insignificant data. Vannamei shrimp infected WFD tended to swim slower and often rose to the surface with body color turned red. Moreover, total organic matter (TOM) increased on week-7 along with the increasing growth of plankton, particularly from the Dinoflagellata group. However, bacterial growth of *Vibrio* sp. on week-7 was insignificant, yet many shrimps were found dead with white feces during that period. The result of analysis indicated that white feces disease was caused by Dinoflagellata besides the main trigger, namely, *Vibrio* sp.

Cite this as: Adam, M. A., Widiastuti, I. M., Ernawati, Yayan, A. Y., Insivitawati, E., Yuliana, Pakaya, R. F., Soegianto, A., & Khumaidi, A. (2022). Analysis of White Feces Disease (WFD) caused by *Vibrio* sp. and Dinoflagellata in Vannamei Shrimp (*Litopenaeus vannamei*) in Brackishwater Culture Pond. *Jurnal Ilmiah Perikanan dan Kelautan*, 14(1):160–166. <http://doi.org/10.20473/jipk.v14i1.26684>

1. Introduction

Shrimp, including vannamei shrimp, is a superior commodity of aquaculture, particularly for export purposes. Vannamei shrimp (*Litopenaues vannamei*) is known as a fast growing shrimp that can be cultured in high stocking density (Aldama-Cano et al., 2018), has high export value and disease resistance (Nur'aini et al., 2019). However, the problem faced by shrimp farmers is mass mortality due to disease attack in shrimp pond (Sun et al., 2013). Various diseases are successfully identified and resulted in great loss, either caused by bacteria (Li et al., 2016) or viruses (Patil et al., 2021), including the White Feces Disease (WFD) in 2012 (Fatmala et al., 2019).

White Feces Disease (WFD) is caused by the attack of *Vibrio* sp. (Longyant et al., 2008). Clinical symptoms shown by shrimp infected with WFD include decreased appetite (Sanguanrut et al., 2018), intestine which turns white or even empty due to lack of food intake (Wang et al., 2020), white feces floating on the water surface (Apitanyasai et al., 2016), and abnormal growth (Tang et al., 2015). Moreover, the disease is also caused by poor aquaculture management (Velmurugan et al., 2015), unhealthy shrimp seed (Aranguren et al., 2017), poor water quality (Mello et al., 2011), and the limited knowledge of shrimp farmers concerning disease that attacks shrimp thus results in slow disease treatment (Rakasiwi & Albastomi, 2017). Shrimp farmers should increase their knowledge about shrimp disease to appropriately prevent and treat the disease (Wahyudi & Fadlil, 2013). WFD is rapidly distributed through water, particularly in ponds with high-stocking density, thus quick treatment is necessary (Rusadi et al., 2019).

However, in several cases, WFD occurs together with the existence of several types of toxic plankton (Ning et al., 2019), such as Dinoflagellata which causes Paralytic Shellfish Poisoning (PSP) disease in shrimp and fish (Adnan, 2014). At high concentrations, Harmful Algal Bloom (HAB) or blooming of toxic phytoplankton may occur (Choirun et al., 2015). Dinoflagellata is identified as toxic species which produces toxins that will enter the food chain of shrimp (Yuliana et al., 2020). In other words, shrimp that consumes the plankton will be dead. The appearance of the disease which possibly attacks vannamei shrimp and the lack of experts who are able to diagnose shrimp disease according to the clinical symptom requires appropriate solution, hence the type of diseases that attack shrimp could be identified effectively and quickly by observing the symptom appears (Adam et al., 2019).

This condition underlined the study on the

occurrence of WFD in vannamei shrimp pond. This study aimed to analyze the appearance of WFD caused by the abundance of *Vibrio* sp. bacteria and Dinoflagellata plankton.

2. Materials and Methods

The study was conducted from December 2020 – March 2021 in the pilot brackish water Pond of PT. Matahari Sakti, Mangunharjo, Central Java. Parameters observed in this study included Total *Vibrio* Count (TVC), clinical symptom, plankton abundance, and water quality. This study survey method washthrough purposive sampling. Sampling location was purposely determined by certain reasons and considerations to collect samples that could represent the area or sample group, allowing researchers to obtain a comprehensive description of the research location. Sampling was done three times in each feeding tray to collect ten shrimps from the pond. Sampling was performed twice a week.

2.1 Total *Vibrio* Count

Bacterial counting was conducted at the end of sampling. Shrimp intestine contents were collected and dissolved in 9 ml of NaCl solvent to be further mixed using vortex until homogenous. Later, 1 ml of sample was cultured on TCBS agar and incubated for 24 hours at 31.4°C (Jayadi et al., 2016).

2.2 Clinical Symptom

Observation of clinical symptom was performed by checking abnormal change that occurred in experimental shrimp. Clinical symptom of shrimp was observed by looking for any changes in symptom that might occur after diet treatment was given, such as appetite, change in body color, change in intestine color, and shrimp behavior. The data obtained were given a score by referring to (Alfiansah & Gardes, 2019). Normal or healthy shrimp is visually active with bright white body color, filled intestinal tract, and non-porous carapace.

2.3 Plankton Abundance

The method applied to count total plankton was the swept area method (Tait, 1972). The calculation was done under the microscope at 10x10 magnification and repeated for three times. Plankton abundance was determined using the formula (APHA, 1998).

$$N = Z \times \frac{X}{Y} \times \frac{1}{V}$$

Description:

N = Individual abundance of phytoplankton (individual/liter)

Z = Individual number of phytoplankton
 X = Volume of filtered water sample (40 ml)
 Y = Volume of 1 drop of water (0.06 ml)
 V = Volume of filtered water (100 l)

2.4 Water Quality

Parameter of water quality observed during the study included temperature, pH, salinity, alkalinity, NO₂, NO₃, NH₄, TOM, PO₄, and N/P ratio. Measurement of water quality was done for 8 weeks of culture.

3. Results and Discussion

3.1 Total *Vibrio* Count (TVC)

The result of the study showed a fluctuating increase in bacterial count on the third week, however it could be controlled and remained stable since the fourth week. Total bacterial count experienced another increase on the eighth week, but total *Vibrio* count remained stable (Figure 1). Total *Vibrio* count of 440 CFU/ml of the total bacterial count (239.875 CFU/ml) (Figure 2).

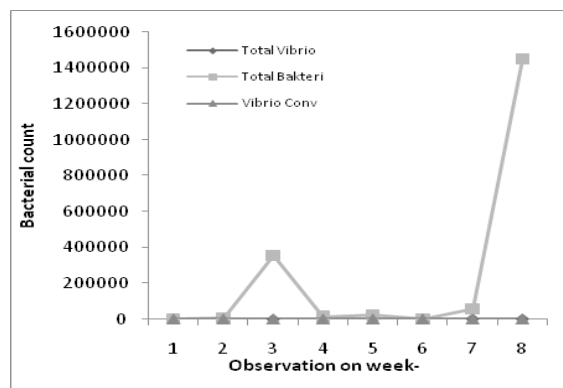


Figure 1. Daily bacterial count in vannamei shrimp culture.

Vibrio bacteria are normally found in waters, yet they are not dangerous if their number is still below the safety limit. Maximum bacterial count threshold for *Vibrio* sp. in water is 10⁴ CFU/ml, while it is 10⁶ CFU/ml in open waters. If bacterial count exceeds this threshold (Sanguanrut *et al.*, 2018), mass mortality of cultured shrimp in the pond may occur (Hatmanti, 2003; Lestari *et al.*, 2018; Roza & Zafran, 1998).

3.2 Clinical Symptoms

The clinical symptom was observed through changes in vannamei shrimp pattern and behavior during culture. Measurement of clinical symptom was done to identify and examine the changes that occurred. Observation conducted on vannamei shrimp infected with WFD

resulted in insignificant data. Vannamei shrimp attacked by WFD in pond tended to swim slower and often rose to the surface with body color that turned red. In 24 hours after the symptom appeared, many shrimps were found dead.

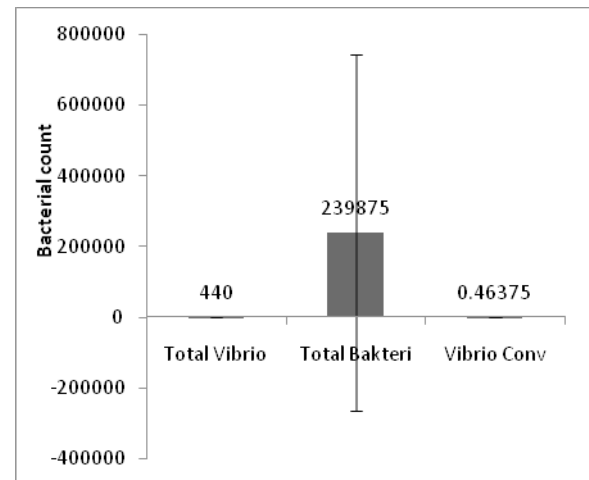


Figure 2. Comparison of bacterial count in vannamei shrimp culture.

3.3 Identification and Plankton Abundance

The result of identification showed plankton diversity in waters which included green algae, blue green algae, Diatom, Euglena, Dinoflagellata, and Protozoa (Figure 4). The study result found three dominant group of plankton, namely Green Algae, Blue Green Algae, and Diatom. However, Green Algae were found to have the highest abundance and diversity (Figure 3).

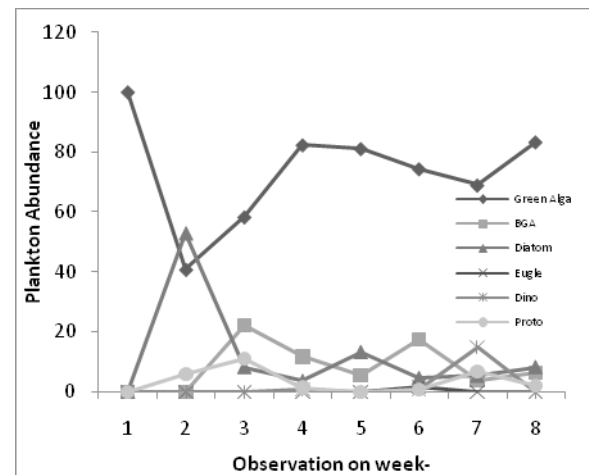


Figure 3. Daily plankton abundance in vannamei shrimp culture.

Green Algae (Chlorophyta) are the main producer in aquatic ecosystem since most (single cell and motile) phytoplankton are member of chlorophyta group which have chlorophyll pigment to effectively perform photosynthesis (Fauziah & Laily, 2015). This result of Green Algae domination showed that the water have

excellent light intensity. Chlorophyta contains higher pigment of chlorophyll a and chlorophyll b compared to carotene and xanthophyll; is a cosmopolitan species; particularly lives in waters with adequate light such as in pond, lake, puddle, and flowing water like river and drain (Siregar & Hermana, 2012). Chlorophyta is also found in semi aquatic environment, such as rocks, damp soil, and damp tree banks (Ariana et al., 2014).

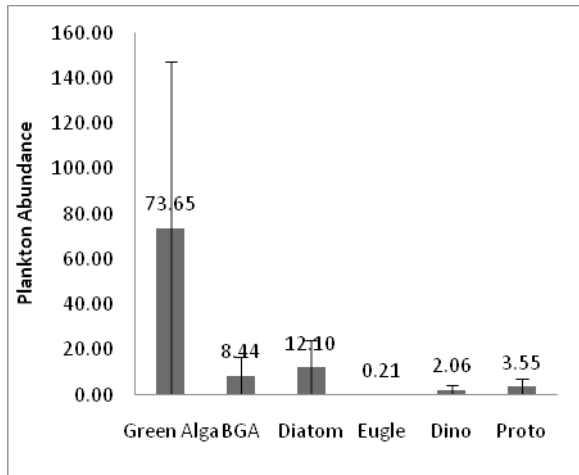


Figure 4. Comparison of plankton abundance in vannamei shrimp culture

3.4 Water Quality

In an aquaculture system, physical and chemical parameters highly affect both organic and inorganic nutrients. The additional treatment applied in the form of lime and fermented probiotics also play important role. The number of plankton increased along with the culture period. Observation of water quality during shrimp culture reflected weekly increase in alkalinity concentration, yet the value was still within the standard for shrimp culture of 100-150 mg/l (Supito et al., 2017). This finding showed that alkalinity is pH buffer in waters. Moreover, (Suwarsih et al., 2016) stated that the increase in alkalinity also affects the nutrient availability which functions as foods for phytoplankton, where increasing alkalinity will result in phosphorus release (Adam & Maftuch, 2015) and increase the availability of carbon for photosynthesis process of phytoplankton.

Result of the study showed an increase in total organic meter (TOM) on week-7 along with the growth of plankton, particularly Dinoflagellata. Moreover, bacterial growth of *Vibrio* sp. on week-7 was found to be insignificant. However, vannamei shrimps were found dead with feces during week-7. This situation indicated that the white feces observed was caused by dinoflagellates. According to Alfiansah and Gardes (2019), population of phytoplankton will increase along with nutrient intake in the pond and duration of

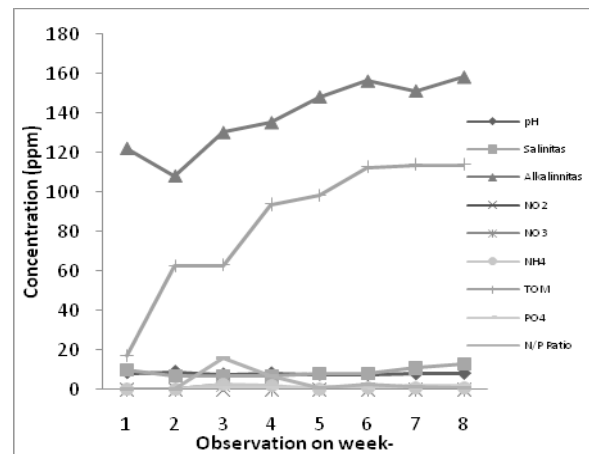


Figure 5. Daily water quality in vannamei shrimp culture.

shrimp culture. Concentration of ammonium and nitrite may cause vannamei shrimp mortality (Sun et al., 2013). *Alteromonas*, *Photobacterium*, *Pseudoalteromonas*, and *Vibrio* are bacteria dominantly found when shrimp disease occurs (Maftuch et al., 2016), either in sample of pond water or in the infected shrimp. Moreover, (Marbun et al., 2019) stated that clinical symptom of shrimp attacked by white feces disease is indicated by change in intestine, such as intestine damage and unfilled intestine content (Aldama-Cano et al., 2018). Furthermore, shrimp are also found to eat less with white pale body color and hepatopancreas damage (Adam et al., 2019). Later, (Nur'aini et al., 2019) said that the pathology of vannamei shrimp infected with WFD indicates the existence of bacterial mass in tubular lumen of hepatopancreas and nodular formation in tubular lumen hepatopancreas caused by high stocking density (Tang et al., 2015). Hence, the growth of bacteria and plankton is related to the increase of total organic matter.

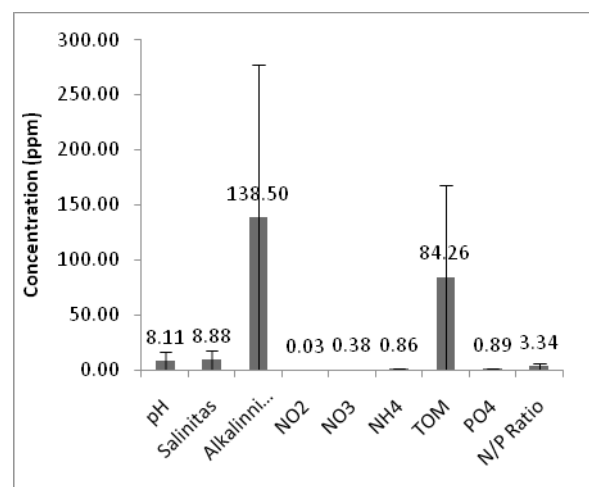


Figure 6. Comparison of water quality in vannamei shrimp culture.

4. Conclusion

Result of analysis indicated that white feces appeared along with the appearance of dinoflagellates which resulted in the disease besides its main trigger which is *Vibrio* spp.

The appearance of the disease which possibly attacks vannamei shrimp and the lack of experts who are able to diagnose shrimp disease according to the clinical symptom requires appropriate solution, hence the type of diseases that attack shrimp could be identified effectively and quickly by observing the symptom appears.

Acknowledgement

This study was funded by the Ministry of Education, Culture, Research and Technology, the Republic of Indonesia.

Authors' Contributions

All authors have contributed to the final manuscript. The contribution of each author as follows, Moh. Awaludin Adam and Ach. Khumaidi; collected the data, drafted and designed the figures. Ira Mei Widiastuti, Ernawati, Achmad Yani, Era Invisitawati, Yuliana, and Rini Fitriyanti Pakaya conceptual ideas and critical revision of the article. All authors discussed the results and contributed to the final manuscript.

Conflict of Interest

The authors declare that they have no competing interests.

Funding Information

This research was partially supported by Universitas Ibrahimy with grant number: 028/AMD-SP2H/LT-MULTI-PDPK/LL7/2021.

References

Adam, M. A., & Maftuch, M. (2015). Evaluasi pengoptimalan instalasi pengolahan air limbah terhadap pencemaran Sungai Wangi di Pasuruan. *Journal of Environmental Engineering and Sustainable Technology*, 2(1):1–5.

Adam, M. A., Maftuch, M., Kilawati, Y., & Risjani, Y. (2019a). Clinical symptoms of gambusia fish (*Gambusia affinis*) after exposure to cadmium absorbed in the gills. *Pollution Research*,

38(March Suppl. Issue):88–93.

Adam, M. A., Maftuch, M., Kilawati, Y., & Risjani, Y. (2019b). The effect of cadmium exposure on the cytoskeleton and morphology of the gill chloride cells in juvenile mosquito fish (*Gambusia affinis*). *Egyptian Journal of Aquatic Research*, 45(4):337–343.

Adnan, Q. (2014). Red Tide. *Oceana*, X(2):65–66.

Aldama-Cano, D. J., Sanguanrut, P., Munkongwongsiri, N., Ibarra-Gómez, J. C., Itsathitphaisarn, O., Vanichviriyakit, R., Flegel, T. W., Sritunyalucksana, K., & Thitamadee, S. (2018). Bioassay for spore polar tube extrusion of shrimp *Enterocytozoon hepatopenaei* (EHP). *Aquaculture*, 490:156–161.

Alfiansah, Y., & Gardes, A. (2019). Manajemen kualitas air dan komunitas bakteri pada tambak udang di Rembang, Indonesia: menuju akuakultur udang yang berkelanjutan. LIPI, Policy Brief #5:20200309. 19.004

APHA. (1998). APHA, A. P. H. A. : Standard methods for the examination of water and wastewater. *American Physical Education Review*, 24(9):481–486.

Apitanyasai, K., Noonin, C., Tassanakajon, A., Söderhäll, I., & Söderhäll, K. (2016). Characterization of a hemocyte homeostasis-associated-like protein (HHAP) in the freshwater crayfish *Pacifastacus leniusculus*. *Fish and Shellfish Immunology*, 58:429–435.

Aranguren, L. F., Han, J. E., & Tang, K. F. J. (2017). *Enterocytozoon hepatopenaei* (EHP) is a risk factor for acute hepatopancreatic necrosis disease (AHPND) and septic hepatopancreatic necrosis (SHPN) in the Pacific white shrimp *Penaeus vannamei*. *Aquaculture*, 471:37–42.

Ariana, D., Samiaji, J., & Nasution, S. (2014). Komposisi jenis dan kelimpahan fitoplankton perairan laut Riau. *Jurnal Online Mahasiswa*, 1–15.

Choirun, A., Sari, S., & Iranawati, F. (2015). Identifikasi fitoplankton spesies Harmfull Algae Bloom (HAB) saat kondisi pasang di Perairan Pesisir Brondong, Lamongan, Jawa Timur. *Jurnal Administrasi dan Kebijakan Kesehatan Indonesia*, 25(2):58–66.

Fatmala, I., Pranggono, H., & Linayati. (2019).

- Identifikasi Bakteri *Vibrio* sp dalam Hepatopankreas udang vannamei (*Litopenaeus vannamei*) pada tambak yang diberi probiotik di Tambak Sampang Tigo Kelurahan Degau Kota Pekalongan. *Jurnal Litbang Kota Pekalongan*, 16:42–48.
- Fauziah, S., & Laily, A. (2015). Identifikasi mikroalga dari Divisi Chlorophyta di Waduk Sumber Air Jaya Dusun Kribet Kecamatan Bululawang Kabupaten Malang. *Bioedukasi: Jurnal Pendidikan Biologi*, 8(1):20.
- Hatmanti, A. (2003). Penyakit bakterial pada budidaya krustasea serta cara penanganannya. *Oseana*, 3(3):1–10.
- Jayadi, M., Prajitno, A., & Maftuch. (2016). The identification of *Vibrio* spp. bacteria from *Litopenaeus vannamei* infected by white feces syndrome. *International Journal of ChemTech Research*, 9(7):448–452.
- Lestari, N., Julyantoro, P., & Suryaningtyas, W. (2018). Uji tantang bakteri *Vibrio harveyi* pada pasca larva udang vaname (*Litopenaeus vannamei*). *Current Trends in Aquatic Science*, 119:112–119.
- Li, S., Li, F., Sun, Z., Zhang, X., & Xiang, J. (2016). Differentially proteomic analysis of the Chinese shrimp at WSSV latent and acute infection stages by iTRAQ approach. *Fish and Shellfish Immunology*, 54:629–638.
- Longyant, S., Rukpratanporn, S., Chaivisuthangkura, P., Suksawad, P., Srisuk, C., Sithigorngul, W., Piyatiratitivorakul, S., & Sithigorngul, P. (2008). Identification of *Vibrio* spp. in vibriosis *Penaeus vannamei* using developed monoclonal antibodies. *Journal of Invertebrate Pathology*, 98(1):63–68.
- Maftuch, Kurniawati, I., Adam, A., & Zamzami, I. (2016). Antibacterial effect of *Gracilaria verrucosa* bioactive on fish pathogenic bacteria. *Egyptian Journal of Aquatic Research*, 42(4):405–410.
- Marbun, J., Harpeni, E., & Wardiyanto, W. (2019). Penanganan penyakit white feces pada udang vaname *Litopenaeus vannamei* menggunakan aplikasi pakan yang dicampur ekstrak lengkuas merah *Alpinia purpurata* k. schum. *Depik Jurnal Ilmu Ilmu Perairan, Pesisir, dan Perikanan*, 8(2):76–86.
- Mello, M. V. C., Aragão, M. E. F. de, Torres-Franklin, M. L. P., Neto, J. M. de O., & Guedes, M. I. F. (2011). Purification of Infectious Myonecrosis Virus (IMNV) in species of marine shrimp *Litopenaeus vannamei* in the State of Ceará. *Journal of Virological Methods*, 177(1):10–14.
- Ning, M., Wei, P., Shen, H., Wan, X., Jin, M., Li, X., Shi, H., Qiao, Y., Jiang, G., Gu, W., Wang, W., Wang, L., & Meng, Q. (2019). Proteomic and metabolomic responses in hepatopancreas of whiteleg shrimp *Litopenaeus vannamei* infected by microsporidian *Enterocytozoon hepatopenaei*. *Fish and Shellfish Immunology*, 87:534–545.
- Nur'aini, Y., Hanggono, B., & Faries, A. (2019). Penanggulangan penyakit berak putih pada udang vaname (*Litopenaeus vannamei*). *Jurnal Perekayasaan Budidaya Air Payau Dan Laut*, 14:108–117.
- Patil, P. K., Geetha, R., Ravisankar, T., Avunje, S., Solanki, H. G., Abraham, T. J., Vinoth, S. P., Jithendran, K. P., Alavandi, S. V., & Vijayan, K. K. (2021). Economic loss due to diseases in Indian shrimp farming with special reference to *Enterocytozoon hepatopenaei* (EHP) and white spot syndrome virus (WSSV). *Aquaculture*, 533:736231.
- Rakasiwi, S., & Albastomi, T. S. (2017). Sistem pakar diagnosa penyakit udang vannamei menggunakan metode forward chaining berbasis web. *Simetris : Jurnal Teknik Mesin, Elektro Dan Ilmu Komputer*, 8(2):647.
- Roza, D., & Zafran. (1998). Pengendalian *Vibrio harvey* secara biologis pada larva udang windu (*Panaeus monodon*) : aplikasi bakteri penghambat. *Jurnal Penelitian Perikanan Indonesia*, 4(2):24–30.
- Rusadi, D., Wardiyanto, & Diantari, R. (2019). Treatment of Vibriosis disease (*Vibrio harveyi*) in vaname shrimp (*Litopenaeus vannamei*, Boone 1931) using *Avicennia alba* leaves extract. *E-Jurnal Rekayasa Dan Teknologi Budidaya Perairan*, 7(2):909.
- Sanguanrut, P., Munkongwongsiri, N., Kongkumnerd, J., Thawonsuwan, J., Thitamadee, S., Boonyawiwat, V., Tanasomwang, V., Flegel, T. W., & Sritunyaluksana, K. (2018). A cohort study of 196 Thai shrimp ponds reveals a complex etiology for early mortality syndrome (EMS). *Aquaculture*, 493:26–36.

- Siregar, B. I. T., & Hermana, J. (2012). Identifikasi dominasi genus alga pada air boezem Morokembrangan sebagai sistem High Rate Algae Pond (HRAP). *Teknik Lingkungan FTSP ITS*, i–xxxiv.
- Sun, Y., Li, F., & Xiang, J. (2013). Analysis on the dynamic changes of the amount of WSSV in Chinese shrimp *Fenneropenaeus chinensis* during infection. *Aquaculture*, 376–379:124–132.
- Supito, Darmawan, A. W., Arief, T., & Iwan, S. (2017). Petunjuk teknis teknik budidaya udang windu (*Penaeus monodon*) pola sederhana melalui penerapan BMPs (Best Management Practices). Jepara: Balai Besar Perikanan Budidaya Air Payau (BBPBAP).
- Suwarsih, Marsoedi, Harahab, N., & Mahmudi, M. (2016). Kondisi kualitas air pada budidaya udang di tambak wilayah pesisir Kecamatan Palang Kabupaten Tuban. Paper presented at *Prosiding Seminar Nasional Kelautan* (pp. 138-143). Madura, Indonesia: University of Trunojoyo Madura.
- Tait, R. (1972). Marine plankton. In R. Tait (Ed.), *Elements of marine ecology. An introductory course.* (pp. 309-312). New York: Springer.
- Tang, K. F. J., Pantoja, C. R., Redman, R. M., Han, J. E., Tran, L. H., & Lightner, D. V. (2015). Development of in situ hybridization and PCR assays for the detection of *Enterocytozoon hepatopenaei* (EHP), a microsporidian parasite infecting penaeid shrimp. *Journal of Invertebrate Pathology*, 130:37–41.
- Velmurugan, S., Palanikumar, P., Velayuthani, P., Donio, M. B. S., Babu, M. M., Lelin, C., Sudhakar, S., & Citarasu, T. (2015). Bacterial white patch disease caused by *Bacillus cereus*, a new emerging disease in semi-intensive culture of *Litopenaeus vannamei*. *Aquaculture*, 444:49–54.
- Wahyudi, M., & Fadlil, A. (2013). Sistem pakar untuk mengidentifikasi penyakit udang galah dengan metode theorem bayes. *Jurnal Sarjana Teknik Informatika*, 1(1):11–20.
- Wang, H., Wan, X., Xie, G., Dong, X., Wang, X., & Huang, J. (2020). Insights into the histopathology and microbiome of Pacific white shrimp, *Penaeus vannamei*, suffering from white feces syndrome. *Aquaculture*, 527:735447.
- Yuliana, Utama, I. G. N. P., & Adam, M. A. (2020). Acute lethal toxicity test of Cd 2+ against gambusia (*Gambusia affinis*) and influence on protease activity. *Samakia: Jurnal Ilmu Perikanan*, 11(1):51–57.