

BIOMECHANIC STUDY OF GRAFT BONE TUNNEL MODEL IN ANTERIOR CRUCIATE LIGAMENT RECONSTRUCTION USING INTRATUNNEL ALLOGENIC BONE MARROW MESENCHYMAL STEM CELLS (BM-MSCs) AND VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF)

Brian Vicky Faridyan¹, Dwikora Novembri Utomo^{1,2}

¹Department of Orthopaedic, Faculty of Medicine, Airlangga University –Dr. Soetomo Hospital, Surabaya, Indonesia.

²Stem Cell Research and Development Center, Universitas Airlangga Surabaya, East Java, Indonesia.

ABSTRACT

Successful anterior cruciate ligament (ACL) reconstruction using tendon graft requires good and rapid integration between the tendon graft and the bone tunnel. The strength of the tendon-bone tunnel graft in the initial phase is very important to facilitate aggressive rehabilitation and as early as possible to support rapid recovery to normal activities. The objective of this study was to determine ultimate tension strength (UTS) on the femoral tendon-bone tunnel graft model after reconstruction of anterior cruciate ligament (ACL) by administering allogenic bone marrow mesenchymal stem cells (BM-MSCs) and vascular endothelial growth factor (VEGF) intratunnel in experimental animals. The design of this research was Post-Test Only Control Group Design using 24 rabbits divided into treatment and control group. Biomechanical evaluation was done at week 3 and 6. Evaluation at week 3 found ultimate tension strength of treatment group significantly higher than control ($p < 0,05$). In the 6th week evaluation, Ultimate tension strength was found that the treatment group significantly higher than the control group ($p < 0,05$). Ultimate tension strength at week 3 did not differ significantly with week 6 ($p > 0,05$). Intravenous administration of BM-MSCs and VEGF on ACL reconstruction increased ultimate tension strength in graft-bone tunnel significantly since week 3. The study of Ferdiansis et al using BM-MSCs and VEGF intraarticular, only showed a significant increase in ultimate tension strength in graft-bone tunnel since week 6. Comparison of this method indicates acceleration in incorporation of tendon graft with bone tunnel on intratunnel method better than invitro intraarticular method.

Keywords : Anterior cruciate ligament, allogenic bone marrow mesenchymal stem cells, vascular endothelial growth factor and biomechanic study.

INTRODUCTION

Anterior Cruciate Ligament (ACL) reconstruction varies considerably in terms of surgical technique, graft source used, and the graft fixation method (Alpert et al., 2008). The main weakness of ACL reconstruction using soft tissue graft is the weak initial relationship

of the tendon-bone interface. The power of graft-bone tunnel in the early phases becomes crucial to facilitate aggressive rehabilitation, so recovery processes including functional exercises, exercise and daily activities can be undertaken soon (Dong et al., 2012). Therefore, the success of ACL reconstruction

using a tendon graft requires good and rapid integration between the tendon graft and the bone tunnel (Dong et al., 2012; Baxter & Bach, 2010; Ferdiansis, 2014).

Bone Marrow-Derived Mesenchymal Stem Cells (BMSCs) are multipotent stem cells and have become an important cell source for engineering purposes in terms of tissue repair and cell therapy (Griffin et al., 2010). Several studies have shown the effects of several polypeptides such as Bone Morphogenetic Proteins (BMPs), and Growth Factors (GFs) such as Transforming Growth Factors (FGFs), Platelet-Derived Growth Factors (PDGF), and Epidermal Growth Factor (EGF) to the activation and regulation of proliferation and differentiation of bone tissue, fibrous tissue, and soft bone tissue (Milano et al., 2008; Dong et al., 2012; Ma et al., 2007; Zelle et al., 2005).

Another problem with post-reconstruction ACL healing process is the occurrence of hypoxia in the tendon tissue autograft. These conditions cause the occurrence of hypocellular and early degeneration conditions

METHODS

This research was a true experimental laboratory study which samples were divided into control group and treatment group. The study design used randomized post test only controlled group design. In this study ACL reconstruction using hamstring autograft tendon which then divided into the control group and treatment as follows:

1. Control group (K1) performed ACL reconstruction with hamstring autograft tendon on graft-tunnel, given

that cause a risk of rupture and elongation in the structure of the tendon graft. Study in Japan, VEGF has been shown to increase vascularization and improve the quality of fibroblast tissue integration between the tendon graft and bone tunnel (Ju et al., 2006; Reinert et al., 2013). Keep in mind also that synovial fluid contains enzymes that cause haematomas as early healing phases are difficult to form (Kaplan & Fu, 2004). Thus the provision of fibrin plugs on the articular side to block the entry of synovial fluid may have a positive effect on bone tunnel integration. In addition, using such plugs allows treatment no longer to be intraarticular, but may be given intratunnel.

Based on that idea, this study was made to determine the role of Bone Marrow-derived Mesenchymal Stem Cells (BMSCs) + Vascular Endothelial Growth Factor (VEGF) intra-tunnel to biomechanical strength and acceleration of post-reconstruction tendon-bone graft integration.

phosphate buffer saline intratunnel, but not given glue seal glue or injection of BMSCs + VEGF, this group will be tested biomechanics at week 3.

2. The control group (K2) performed ACL reconstruction with hamstring autograft tendon on the graft-tunnel, given intravenous saline phosphate buffer, but not given glue-seal fibrin and BMSCs + VEGF injection, the group will be tested biomechanically at week 6.

3. The second treatment group (P1) after ACL reconstruction with hamstring autograft tendon on the graft-tunnel, followed by fibrin glue seal at the end of the intraarticular tunnel, then injected BMSCs + VEGF on the intratunnel, the group will be tested biomechanically at week3.
4. The second treatment group (P2), after ACL reconstruction with hamstring autograft tendon on the graft-tunnel, followed by fibrin glue seal at the end of the intraarticular tunnel, then injected BMSCs + VEGF on the

intratunnel, the group will be tested biomechanically at week to-6.

The results obtained were evaluated biomechanically at third week and sixth weeks after treatment with different sample units. The sample size of this study is 6, with the calculation of the number of test groups is 4 then the required sample size is 24 samples. The research was conducted at Stem Cell Research and Development Center of Airlangga University Surabaya. The study was conducted for 8 weeks, the first 2 weeks for culture process from BM-MSCs and 6 weeks later for treatment and evaluation process.

RESULTS

The ultimate tension strength evaluation showed that the mean of ultimate tension

strength of control group and treatment at week 3 was 0.840 ± 0.3089 N and 4.340 ± 1.0188 N according to table 1 below.

Table 1. Evaluation Ultimate Tension Strength (N) at week 3

Group	No. Sample	Ultimate Tension Strength (N)	Explanation	$\bar{x} \pm SD$
Control	C3.1	0,93	Pullout	$1,025 \pm 0,1415$
	C3.2	1,12	Ruptur	
	C3.3	1,17	Pullout	
	C3.4	0,88	Pullout	
Treatment	T3.1	1,25	Ruptur	$1,378 \pm 0,1011$
	T3.2	1,40	Ruptur	
	T3.3	1,32	Ruptur	
	T3.4	1,52	Pullout	
	T3.5	1,40	Ruptur	

From table 2 it was found that after 6 weeks evaluation, the mean ultimate tension strength

in the control group was 1.025 ± 0.1415 N and 1.378 ± 0.1011 N in the treatment group.

Table 2. Evaluation Ultimate Tension Strength (N) at week 6

Group	No. Sample	Ultimate Tension Strength (N)	Explanation	$\bar{x} \pm SD$
Control	C6.1	1,25	Pullout	$1,280 \pm 0,0474$
	C6.2	1,28	Pullout	
	C6.3	1,34	Ruptur	
	C6.4	1,22	Pullout	
	C6.5	1,31	Ruptur	

Treatment	T6.1	1,29	Ruptur	1,4667 ± 0,1257
	T6.2	1,59	Pullout	
	T6.3	1,43	Ruptur	
	T6.4	1,36	Ruptur	
	T6.5	1,55	Ruptur	
	T6.6	1,58	Ruptur	

Table 3. Comparison of Ultimate Tension Strength of Control and Treatment Group on Week 3

Group	N	$\bar{x} \pm SD$	p
Control	4	1.025 ± 0.1415	0,003
Treatment	5	1.378 ± 0.1011	

Table 3 shows that in the treatment group using BM-MSCs intratunnel had ultimate tension

strength significantly greater than control group ($p < 0.05$).

Table 4. Comparison of Ultimate Tension Strength of Control and Treatment Groups at Week 6

Group	N	$\bar{x} \pm SD$	p
Control	5	1.280 ± 0.0474	0,012
Treatment	6	1.467 ± 0.1256	

From table 4 found in the treatment group given intratunnel BM-MCs, after the sixth week the ultimate tension strength was greater

than the control group and statistically significant ($p < 0.05$).

Table 5. Comparison of Ultimate Tension Strength Treatment Groups at 3 weeks and 6 weeks

Group	N	$\bar{x} \pm SD$	p
3 weeks	5	1.378 ± 0.1011	0,236
6 weeks	6	1.467 ± 0.12	

When compared to the evaluation of ultimate tension strength of treatment group at week 3 and week 6, it can be seen from Table 4.6 that

there was no significant difference between the two groups ($p > 0.05$).

DISCUSSION

ACL reconstruction surgery aims to restore knee biomechanical stability. The power of graft-bone tunnel in the early phases becomes crucial to facilitate aggressive rehabilitation, so recovery processes including functional exercises, exercise and daily activities can be undertaken soon (Dong et al., 2012). Therefore, the success of ACL reconstruction using tendon graft requires good and rapid integration between the tendon graft and the bone tunnel (Griffin et al., 2010).

The process of tissue formation between the tendon and bone is the most important stage at the beginning of the healing process. Therefore, it is important to determine how to heal the graft-tunnel as early as possible. In the healing process of the tendon-bone, a fibrovascular scar layer develops between the tendon and bone in the graft-tunnel interface. This layer will eventually arrange the fibers that are perpendicular to each other to resemble Sharpey's fiber. The presence and amount of these fibers is directly proportional to ultimate

tension strength.

Bone Marrow-Derived Mesenchymal Stem Cells (BMSCs) are multipotent stem cells and have become an important cell source for engineering purposes in terms of tissue repair and cell therapy (Griffin et al., 2010). Studies have shown that a bone tunnel with increasing amount of BMSCs can increase tendon-to-bone healing in rabbit models through the formation of perpendicular collagen fibers connecting the tendon to bone (resembling Sharpey's fibers) and an increase in the number of cartilage-like cells proliferation by week four (Dong et al., 2012; Milano et al., 2008; Ma et al., 2007; Kohno et al., 2007; HW et al., 2004). Given growth factor VEGF has been shown to increase vascularization and improve the quality of fibroblast tissue integration between the tendon graft and bone tunnel (Reinert et al., 2013).

This study was a true experimental laboratory study evaluating ultimate tension strength biomechanically on the ACL reconstruction tendon-bone tunnel graft treated with BM-MSCs and VEGF intratunnel by using glue seal fibrin to keep BM-MSCs and VEGF in the bone tunnel. Observations were made at week 3 and week 6 according to an evaluation performed by Tomita et al in 2001 (Tomita et al., 2001). The relative hypoxic conditions of the tendon graft result in tissue degradation. In this study the administration of BM-MSCs and VEGF is expected to promote neovascularization, fibroblast proliferation, collagen synthesis in bone tunnel (Deehan & Cawston, 2005).

The results of the comparative analysis using independent T test in this study showed significant differences in ultimate tension strength between the ACL reconstruction treatment group who were given BM-MSCs and VEGF intratunnel compared with the control group in both 3 weeks observation ($p < 0.05$) and week observation to 6 ($p < 0.05$). However, there was no significant difference between the 3rd week observation group and the 6th week observation ($p > 0.05$), although the results at week 6 (1.467 ± 0.12) showed higher ultimate tension strength compared to week 3 (1.378 ± 0.10).

The Ferdiansis et al study on ACL reconstructed models treated by BM-MSCs + VEGF intra-articular showed no significant difference between the 3rd week treatment with new controls with significant difference after week 6 when compared with control ($p < 0.05$) (Ferdiansis, 2014). While in this study, significant differences with controls can be demonstrated since week 3. This indirectly suggests that the intravenous method of administering BM-MSCs + VEGF will provide an acceleration in the integration of the tendon-bone tunnel graft when compared with intraarticular administration (Ferdiansis, 2014).

The results of this study is in accordance with Kanazawa research, which at week 2 and 4 the expression of type 1 collagen with chondroid coating was formed (Kanazawa et al., 2012). Where the tensile strength of the bone tunnel was aligned with the number of collagen type 1 and the chondroid layer.

CONCLUSION

In the biomechanics test on the tendo-bone tunnel graft model after anterior cruciate ligament reconstruction with intravenous administration of BM-MSCs and VEGF, it can be concluded as follows:

1. Ultimate tensile strength of 3rd week observational group treated by BM-MSCs and VEGF in intratular was significantly higher than control group.
2. The ultimate tensile strength of the 6th

week observational group treated by BM-MSCs and VEGF by intratular was significantly higher than the control group.

3. There was no significant difference in ultimate tensile strength in week 3 and week 6 observation group.
4. This ultimate difference in tensile strength illustrates indirectly to the integration of the tendon-bone tunnel, ie, there is a significant difference in controls since the 3rd week after ACL reconstruction.

REFERENCES

- Alpert, J.M., Brush-Joseph, C.A., Bach, J. & Bernard, R., 2008. Patellar Tendon Autograft for Anterior Cruciate Ligament Reconstruction. In *Surgical Techniques of The Shoulder, Elbow, and The Knee in Sports Medicine*. Philadelphia, PA, USA: Elseviers Saunders.
- Amiel, D., Frank, C. & Harwood, F., 1984. Tendons and ligaments: a morphological and biochemical comparison. *J Orthop Res*, 1, pp.257–65.
- Anderson, A.F., Dome, D.C. & Gautam, S., 2001. Correlation of anthropometric measurements, strength, anterior cruciate ligament size, and intercondylar notch characteristics to sex differences in anterior cruciate ligament tears. *Am J Sports Med*, (29), pp.58–63.
- Bach, J.M., Hull, M.L. & Patterson, H.A., 1997. Direct measurement of strain in the posterolateral bundle of the anterior cruciate ligament. *J.Biomech*, 30(3), pp.281-83.
- Baxter, F.R. & Bach, J.S., 2010. Augmentation of Bone Tunnel Healing in Anterior Cruciate Ligament Grafts: Application of Calcium Phosphates and Other Materials. *Journal of Tissue Engineering*.
- Chandler, J.W. & Creighton, R.A., 2008. Patellar Tendon Allograft for Anterior Cruciate Ligament Reconstruction. In *Surgical Techniques of The Shoulders, Elbow, and Knee in Sport Medicine*. Philadelphia, PA, USA: Elseviers Saunders.
- Colombet, P., Robinson, J. & Christel, P.I., 2006. Morphology of anterior cruciate ligament attachments for anatomic reconstruction: a cadaveric dissection and radiographic study. *Arthroscopy*, 22, pp.984–92.
- Da Silva, E.M. & Albano, M.B., 2013. Knee Ligament Injuries:

- Biomechanics Comparative Study of Two Suture Technique in Tendon Analysis. *Rev Bras Ortop*, 48(1), pp.80-86.
- Deehan, D.J. & Cawston, T.E., 2005. The Biology of Integration of Anterior Cruciate Ligament. *British Editorial Society of Bone and Joint Surgery*, 87(8), pp.889-95.
- Dong, Y. et al., 2012. Enhancement of Tendon-Bone Healing for Anterior Cruciate Ligament (ACL) Reconstruction Using Bone Marrow-Derived Mesenchymal Stem Cells Infected with BMP-2. *International Journal of Molecular Sciences*, 13, pp.13605-20.
- Duthon, V.B., Barea, C. & Abrassart, S., 2006. Anatomy of the anterior cruciate ligament. *Knee Surg Sports Traumatol Arthrosc*, 14, pp.204–13.
- Feng, L., Jia, H. & Yu, C., 2007. ACL reconstruction in a rabbit model using irradiated Achilles allograft seeded with mesenchymal stem cells or PDGF-B gene-transfected mesenchymal stem cells. *Knee Surgery Sport Traumatology Arthroscopy*, Knee Surgery Sport Traumatology Arthroscopy, p.1219027.
- Ferdiansis, e.a., 2014. Effect of intraarticular Bone Marrow Mesenchymal Stem Cells and Vascular Endothelial Growth Factor on Graft-Tunnel Integration after ACL Reconstruction., 2014. National Indonesian Orthopaedic Congress.
- Freeman, A., 2001. How the Knees Move. *Current Orthopaedics*, 15, pp.444 - 450.
- Fukubayashi, T., Torzilli, P.A., Sherman, M.F. & Warren, R.F., 1982. An in vitro biomechanical evaluation of anterior-posterior motion of the knee. Tibial displacement, rotation, and torque. *J. Bone Joint Surg Am.*, 64(258-264), pp.258-64.
- Gotter, D. & Schmidt-Wiethoff, 2007. Biomechanics of the Anterior Cruciate Ligament and Implications for Surgical Reconstruction. In *Strategies Trauma Limb Reconstr.* pp.1-12.
- Griffin, M.D., Ritter, T. & Mahon, B.P., 2010. Immunological Aspects of Allogeneic Mesenchymal Stem Cells Therapies. *Human gene therapy*, 21, pp.1641-55.
- Harner, C.D., Baek, G.H. & Vogrin, T.M., 1999. Quantitative analysis of anterior cruciate ligament insertions. *Arthroscopy*, 15, pp.741–49.
- Hays, P. et al., 2008. The Role of Macrophages in Early Healing of a Tendon Graft in a Bone Tunnel. *Journal Bone Joint Surgery America*, 90-A(3), pp.565-75.
- HW, O., JC, G. & EH, L., 2004. Use of bone marrow stromal cells for tendon graft-to-bone healing: histological and immunohistochemical studies in a rabbit model. *Am J Sports Med*, 32, pp.321-27.
- Jones, C.D. & Grimshaw, P.N., 2011. The Biomechanics of the Anterior Cruciate

- Ligament and Its Reconstruction. In *Theoretical Biomechanics*. pp.361–381.
- Ju, Y.J. et al., 2006. Effects of Local Administration of Vascular Endothelial Growth Factor on Properties of the in Situ Frozen-Thawed Anterior Cruciate Ligament in Rabbits. *The American Journal of Sports Medicine*, 34(1), pp.84-89.
- Kanaya, A. et al., 2007. Intra-articular Injection of Mesenchymal Stromal Cells in Partially Torn Anterior Cruciate Ligaments in a Rat Model. *Arthroscopy: The Journal of Arthroscopic and Related Surgery*, 23(6), pp.610-17.
- Kanazawa, T., Soejima, T. & Tabuchi, K., 2012. Histological analysis on the tendon-to-bone healing utilizing bone marrow-derived MSCs in ACL reconstruction model without a bone tunnel. In *ORS 2012 Annual Meeting*, 2012.
- Kaplan, L.D. & Fu, F., 2004. Clinical Applications of Orthopedic Tissue Engineering: Ligaments and Tendons. In A.C. VM Goldberg, ed. *Orthopedic Tissue Engineering Basic Science and Practice*. New York, USA: Marcel Dekker Inc.
- Kohno, T. et al., 2007. Immunohistochemical demonstration of growth factors at the tendon-bone interface in anterior cruciate ligament reconstruction using a rabbit model. *J. Orthop. Sci.*, 12, pp.67-73.
- Krosshaug, T. & Slauterbeck, J.R., 2006. Biomechanical Analysis of Anterior Cruciate Ligament Injury Mechanisms: Three-Dimensional Motion Reconstruction from Video Sequences. *Scandinavian Journal of Medicine and Science in Sports*.
- Lane, J.G., Kaufman, K., Rangger, C. & Daniel, D.M., 1994. The anterior cruciate ligament in controlling axial rotation. An evaluation of its effect. *Am. J. Sports Med.*, 22(2), pp.289-93.
- L'Insalata, J.C., Klatt, B. & Fu, F.H., 1997. Tunnel expansion following anterior cruciate ligament reconstruction: A comparison of hamstring and patellar tendon autograft. *Knee Surg Sports Traumatol Arthrosc*, 5, pp.234-38.
- Lipke, J.M., Janecki, C.J. & Nelson, C.L., 1981. The role of incompetence of the anterior cruciate and lateral ligaments in anterolateral and anteromedial instability. A biomechanical study of cadaver knees. *J Bone Joint Surg Am*, 63(954-960).
- Ma, C.B. et al., 2007. Bone morphogenetic proteins-signaling plays a role in tendon-to-bone healing: A study of rhBMP-2 and noggin. *Am. J. Sports Med.*, 35, pp.597–604.
- Mall, N.A., Van Thiel, G.S., Bedi, A. & Cole, B.J., 2013. *Graft Selection in Anterior Cruciate Ligament Reconstruction*. [Online] Available at: <http://www.rockfordortho.com/wp-content/uploads/2012/07/Graft-Selection-in-ACL-Surgery.pdf>

- [Accessed 11 January 2015]
- McCarty, L.P., 2005. Anatomy, Biology, and Biomechanics of Patellar Tendon Autograft Anterior Cruciate Ligament Reconstruction. *Techniques in Ortop.*
- Milano, G., Deriu, L. & Fabbriani, C., 2008. Graft-Tunnel Healing. In C. Brown et al., eds. *The Anterior Cruciate Ligament: Reconstruction and Basic Science*. Philadelphia, PA, United State of America: SAUNDERS Elsevier.
- Odensten, M. & Gillquist, J., 1985. Functional anatomy of the anterior cruciate ligament and a rationale for reconstruction. *J Bone Joint Surg Am*, 67, pp.257–62.
- Oe, K., Kushida, T. & Okamoto, N., 2011. New strategies for anterior cruciate ligament partial rupture using bone marrow transplantation in rats. *StemCells Development*, 20, pp.671-79.
- Petersen, W. & Tillmann, B., 2002. Anatomy and function of the anterior cruciate ligament. *Orthopäde*, 31, pp.710–18.
- Pinczewski, L., Clingeleffer, A., Otto, D. & Bonar, F., 1997. Integration of Hamstring Tendon Graft With Bone in Reconstruction of the Anterior Cruciate Ligament. *Arthroscopy: The Journal of Arthroscopic and Related Surgery*, 13(5), pp.641-43.
- Reinert, R.B. et al., 2013. Vascular endothelial growth factor- α and islet vascularization are necessary in developing, but not adult, pancreatic islets. *PubMed.gov Diabetes.*, 62(12), pp.4154-64.
- Rodeo, S.A. et al., 1993. Tendon-healing in a bone tunnel. A biomechanical and histological study in the dog. *Journal Bone Joint Surgery America*, 72(12), pp.1795-803.
- Sakane, M., Fox, R.J. & Woo, S.L., 1997. In situ forces in the anterior cruciate ligament and its bundles in response to anterior tibial loads. *J Orthop Res*, 15, pp.285–93.
- Song, E.K. et al., 2004. Failure of Osteointegration of Hamstring Tendon Autograft After Anterior Cruciate Ligament Reconstruction. *Arthroscopy: The Journal of Arthroscopic and Related Surgery*, 20(4), pp.424-28.
- Tomita, F. et al., 2001. Comparisons of intraosseous graft healing between the doubled flexor tendon graft and the bone-patellar tendon-bone graft in anterior cruciate ligament reconstruction. *Arthroscopy*, 17(5).
- Weiler A et al., 2002. Tendon Healing in a Bone Tunnel. Part I: Biomechanical Results After Biodegradable Interference Fit Fixation in a Model of Anterior Cruciate Ligament Reconstruction in Sheep. *Arthroscopy: The Journal of Arthroscopic and Related Surgery*, 18(2), pp.113-23.
- Weiler, A. et al., 2002. Tendon Healing in a Bone Tunnel Part II: Histologic Analysis After Biodegradable Interference Fit Fixation in a Model of

Anterior Cruciate Ligament
Reconstruction in Sheep. *Arthroscopy:
The Journal of Arthroscopic and
Related Surgery*, 72(12), pp.113-23.

Zelle, B.A. et al., 2005. Biological

Consideration of Tendon Graft
Incorporation Within the Bone
Tunnel. *Elsevier*, Operative Technique
in Orthopaedics.